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1a1 Growth Factor Upregulates the Expression of Nitric Oxide Synthase in Native and Cultured Endothelial Cells

W. B. Schen-Aureth, R. Busse, Zentrum fuer Physiologie, Frankfurt am

growth factor (VEGF) has been shown to accelerate re-endothelialization of infarcting in balloon-injured arteries. Since endothelium-derived nitric oxide promotes smooth muscle growth, we tested whether VEGF enhances nitric oxide synthase activity. Experiments were performed with primary cultures of human aortic endothelial cells (HAEV) and with endothelium-isolated rat aortic rings. Nitric oxide production was assessed by RT-PCR and Western blot analysis, and NOS II mRNA levels were determined by Northern blot analysis. VEGF treatment increased NOS II mRNA levels in HAEV (100 ng/ml) for 48 h to an increase in NOS II mRNA of 2.2 ± 0.5 fold, whereas that induced by VEGF in aortic rings was unaffected. VEGF treatment increased in a concentration-dependent manner NOS II mRNA levels within 2 h which remained elevated for the next 48 h. The effect of VEGF on NOS II mRNA was abolished by protein tyrosine kinase and extracellular signal-regulated kinase (ERK) and was not prevented by the inhibitor of transcription factor NF-κB. Aortic rings to VEGF for 7 h accumulated endothelium-dependent nitric oxide whereas those to SNP were unaffected. Increased levels of NOS II are found in VEGF-treated aortic rings. These findings indicate that VEGF expression in native and cultured endothelial cells. This effect is mediated by a kinase-dependent pathway(s) and seems to be due to the protective role of nitric oxide in endothelial nitric oxide synthase activity.

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Inhibition of Pin, A Protein Inhibitor of Neuronal Nitric Oxide Synthase

Michael T. Greenwood, Oussama El-Divan, Yang Guo, McGill University,

synthase (nNOS) has been localized to the inner membrane of skeletal muscle myofibrils. nNOS is associated to the cytoskeleton and has been proposed to be involved in the regulation of muscle contraction. A protein inhibitor of nNOS (PIN) which specifically inhibits nNOS dimerization has recently been cloned. The 89 residue PIN, regulates the biochemical activity of nNOS. We have, therefore, examined, the distribution and regulation of PIN transcript in skeletal muscle myofibrils. The distribution and regulation of PIN transcript in skeletal muscle myofibrils was examined in skeletal muscle myofibrils. PIN expression has been shown to be increased in skeletal muscle of normal rats but the highest levels of PIN were observed in skeletal muscle of rats with the highest levels of PIN in the skeletal muscle which has the lowest nNOS expression. In the skeletal muscle of PIN transgenic mice, PIN expression was elevated with significant developmental regulation. In embryonic rats (20 mg/kg E.coli), PIN and nNOS expressions were elevated in skeletal muscle within 12 hours of birth. We further examined the levels of PIN and nNOS in the skeletal muscle of growing myofibrils. Significant PIN mRNA expression was detected. High PIN expression was maintained following the induction of myofibril fusion and in the mature state indicates that PIN is expressed in various skeletal muscles in vivo and its expression correlates with that of nNOS. Moreover PIN mRNA is in pathological conditions such as skeletal myopathy.

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Interleukin-1 and Nuclear Factor-κB During S Transcription

Zaragoza, Audrey, McMillan, Charles J, Loveston, John, Houston,

endothelial nitric oxide synthase (eNOS) is expressed in a variety of inflammatory disorders in the endothelium. eNOS expression is regulated at the transcriptional level. We have examined the transcriptional regulation of the eNOS gene. Interleukin-1 (IL-1) and IL-6 activate NF-κB and IRF-1 in macrophages, and these transcription factors bind to the κB and IRF-1 sites in the eNOS promoter to induce eNOS expression. We have examined the interaction between NF-κB and IRF-1 and NF-κB and IRF-1. Co-immunoprecipitation experiments show that IRF-1 and NF-κB interact when binding to DNA. The proteins bound to a κB site include not only NF-κB but also IRF-1. Similarly, the proteins bound to an IRF-1 site include not only IRF-1 but also NF-κB. To explore the functional consequences of the interaction between NF-κB and IRF-1, we examined their ability to affect the structure of the eNOS promoter. We found that the interaction between NF-κB and IRF-1 affects the structure of the eNOS promoter at the site of binding by

Aerobic Nitric Oxide Synthase Gene Transfer in Acute Hypoxic Pulmonary Hypertension

Werner Budts, Carolee Unit, Leuven Belgium; Zengshan Nong, VIB, Leuven Belgium; Natascha van Pelt, Carolee Unit, Leuven Belgium; Rick Lyons, Univ of New Mexico, Albuquerque, NM; Robert D Gerard, VIB, Leuven Belgium; Stefan Janssens, Carolee Unit, Leuven Belgium

Nitric oxide (NO), a vasodilator involved in the regulation of pulmonary vascular tone, is synthesized by a class of enzymes, NO synthases (NOS). We have previously shown that adenovirus-mediated overexpression of the calcium-dependent type II NOS in rat lungs reduces acute hypoxic pulmonary vasoconstriction. To quantify the effect and duration of NO production following NOS gene transfer, we measured exhaled NO by chemoluminescence in rats infected with adenovirus expressing the calcium-independent type II NOS (4x10⁶ pfu/ml, n=7), type II NOS (n=6), or control virus expressing no transgene (AdRRS, n=7). Exhaled NO was increased in NOS II-infected rats compared to NOS II-infected rats at 24 h (55 ppb vs 47 ppb, 4 d (52 ppb vs 36 ppb), and 7 d (31 ppb vs 22 ppb), but no longer at 10 d. The levels of NO in AdRRS-infected rats were significantly lower at all time points (19±8 ppb at 24 h, 6±3 ppb at 4 d and 7±1 ppb at 7 d). To investigate whether increased pulmonary NO production after NOS II gene transfer was associated with greater inhibition of hypoxic pulmonary vasoconstriction, basal pulmonary artery pressure (PAP, mmHg) was measured during acute hypoxia (FIO₂ 0.12, 25 mmHg) in rats 4 d after infection with NOS II (n=7), NOS II (n=6), or control virus (n=6). Acute hypoxia increased PAP from 19±4 to 21±5 mmHg in NOS II-infected rats compared to 23±2 mmHg in NOS II-infected and 25±2 mmHg in control virus-infected rats with no significant effect on systemic blood pressure. Thus, pulmonary NOS II gene transfer significantly increases pulmonary exhaled NO production for at least 7 days and is associated with a greater inhibition of acute hypoxic pulmonary vasoconstriction. Single intrapulmonary NOS II gene transfer may be a promising therapy for pulmonary hypertension.

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A Murine Model of Accelerated Diabetic Atherosclerosis: Suppression By Soluble Receptor For Advanced Glycation Endproducts

Lee Park, Kathleen G Ramon, Kenneth J Lee, Yan Li, Michael D Ginsberg, Luis Ferran Jr, David M Stern, Ann Marie Schmidt, Columbia University, New York, NY

Multiple studies suggest that lipid-independent mechanisms contribute to the development of cardiovascular disease in diabetes. Under conditions of sustained hyperglycemia, nonenzymatic glycation and oxidation of proteins and lipids results in the irreversible formation of Advanced Glycation Endproducts (AGEs) which accumulate in diabetic plasma and tissues. AGEs interact with cellular receptors such as RAGE, the Receptor for AGEs, and induce vascular cell dysfunction. The extracellular portion of RAGE (one V-type and two C-type immunoglobulin domains) is a soluble fragment (sRAGE) which we postulate may bind AGEs and block their interaction with their cell-surface receptor. We previously demonstrated a 3.7-fold decrease in atherosclerotic lesion area in streptozotocin-treated apolipoprotein E deficient mice in controls, with enhanced accumulation of AGEs and increased expression of RAGE in the vasculature by immunohistochemistry. To test if blockade of AGE-RAGE would suppress accelerated atherosclerosis, diabetic and E deficient mice were treated for six weeks with sRAGE (200 μg/day, intraperitoneally) or equivalent mouse serum albumin (400 μg/day MSA). Mean lesion area was decreased 1.8-fold (p=0.016) in mice treated with sRAGE (150.06±18.549 μm²) vs MSA (271.00±16.721 μm²). No difference was observed at a low dosage of 30 μg/day. Serial serum glucose and HbA_{1c} levels revealed persistent hyperglycemia in both groups. There were no differences in levels of total cholesterol and triglycerides, and FPLC analysis yielded identical lipid profiles. Taken together, these data suggest that enhanced AGE-RAGE interaction likely plays a critical role in the pathogenesis of accelerated atherosclerosis in diabetes.

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Essential Role of Endothelial Nitric Oxide Synthase in Angiogenesis in Vivo

Toyoko Muramatsu, Takahito Asahara, Marcy Silver, Monette Kearney, Meredith Magner, Jiong Yang, Donghua Chen, Donglin Chen, James F Symes, St Elizabeth's Medical Center, Boston, MA; Paul L Huang, Massachusetts General Hospital, Boston, MA; Jeffrey M Lerner, St Elizabeth's Medical Center, Boston, MA

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